## Determinand:

Haloacetic Acids and Dalapon

## Matrix:

Sample Types: Treated and Raw waters.

## Principle of Method:

The method is a direct aqueous injection procedure. Samples are analysed by high performance liquid chromatography using a triple quadrupole mass spectrometer as a detector. The aqueous sample is injected and the organic compounds are separated and then identified and quantified with mass spectrometric detection (MSD) in selected reaction monitoring (SRM) mode. Quantitation is based on an internal standardisation procedure.

## Sampling and Sample Preparation:

Sampling, samples should be collected in 250 mL coloured glass bottles with PTFE lined screw caps containing $250 \mu \mathrm{~L}$ of $10 \% \mathrm{w} / \mathrm{v}$ ammonium chloride solution as preservative, i.e. STL430.
Storage - samples should be analysed as soon as possible after collection. When this is not possible, they are stored under refrigeration at 1-5DC until analysis can begin. The maximum permissible storage time prior to analysis is given below:

| Determinand | Maximum period of <br> analyte stability prior <br> to any extraction step <br> (days) | Maximum period of <br> analyte stability after to <br> any extraction step <br> (days) | Data is quoted from BS <br> EN ISO 5667-3: 2003 <br> ["ISO"J or ALS in-house <br> data ["ALS-AS IHD"] |
| :--- | :---: | :---: | :---: |
| MCAA | 10 | $\mathrm{~N} / \mathrm{A}$ | ALS IHD |
| DCAA | 14 | $\mathrm{~N} / \mathrm{A}$ | ALS IHD |
| TCAA | 21 | $\mathrm{~N} / \mathrm{A}$ | ALS IHD |
| MBAA | 14 | $\mathrm{~N} / \mathrm{A}$ | ALS IHD |
| DBAA | 14 | $\mathrm{~N} / \mathrm{A}$ | ALS IHD |
| TBAA | 21 | $\mathrm{~N} / \mathrm{A}$ | ALS IHD |
| BCAA | 14 | $\mathrm{~N} / \mathrm{A}$ | ALS IHD |
| BDCAA | 21 | $\mathrm{~N} / \mathrm{A}$ | ALS IHD |
| DBCAA | 21 | $\mathrm{~N} / \mathrm{A}$ | ALS IHD |
| Dalapon | 21 | $\mathrm{~N} / \mathrm{A}$ | ALS IHD |

## Interferences

HPLC-QQQ is an extremely selective technique and interferences should only be encountered very rarely. Any interfering compounds would have to display the identical SRM transition at the same retention time; this is extremely unlikely in potable water samples. However, any compound, which passes through the extraction procedure, and has a similar liquid chromatographic retention time and mass spectrometric properties to the compound of interest, will cause interference. Samples containing high humic or fulvic loading have been demonstrated to not cause significant ion suppression for the compounds.

## Performance of Method:

## Range of Application:

LOQ to $150 \mu \mathrm{~g} \mathrm{~L}^{-1}$
Samples producing results above this range should be diluted and re-extracted.

## Limit of Quantification:

| Determinand | Method Limit of <br> Quantification ( $\boldsymbol{\mu g ~ L -}$ |
| :--- | :---: |
| Monochloroacetic acid (MCAA) | 1.3 |
| Dichloroacetic acid (DCAA) | 1.2 |
| Trichloroacetic acid (TCAA) | 1.7 |
| Monobromoacetic acid (MBAA) | 1.2 |
| Dibromoacetic acid (DBAA) | 1.6 |
| Tribromoacetic acid (TBAA) | 1.2 |
| Bromochloroacetic acid (BCAA) | 1.0 |
| Bromodichloroacetic acid (BDCAA) | 1.3 |
| Dibromochloroacetic acid (DBCAA) | 1.5 |
| Dalapon | 1.2 |

Recoveries of Compounds, Bias and Uncertainty of measurement:

| Determinand | Uncertainty of Measurement (UoM) (\%) | Direct Standards |  |  |  | Elvington Treated Water <br> PCV Spike |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  | Low Standard |  | High Standard |  |  |  |
|  |  | Recovery | SD | Recovery | SD | Recovery | SD |
| MCAA | 10.554 | 98.39\% $\pm 1.50 \%$ | 3.20\% | 100.33\% $\pm 1.41 \%$ | 3.10\% | 101.18\% $\pm 1.72 \%$ | 3.53\% |
| DCAA | 21.971 | 102.06\% $\pm 2.16 \%$ | 4.79\% | 100.70\% $\pm 2.43 \%$ | 4.86\% | 110.32\% $\pm 2.70 \%$ | 4.85\% |
| TCAA | 9.695 | 99.34\% $\pm 1.50 \%$ | 3.35\% | 100.77\% $\pm 1.36 \%$ | 2.73\% | 101.39\% $\pm 1.45 \%$ | 2.72\% |
| MBAA | 10.865 | 100.91\% $\pm 1.72 \%$ | 3.79\% | 101.60\% $\pm 1.59 \%$ | 3.46\% | 101.71\% $\pm 1.32 \%$ | 2.87\% |
| DBAA | 17.824 | 100.76\% $\pm 1.78 \%$ | 3.69\% | 101.41\% $\pm 2.14 \%$ | 4.47\% | 106.17\% $\pm 2.36 \%$ | 4.40\% |
| TBAA | 17.343 | 106.43\% $\pm 2.17 \%$ | 4.03\% | 100.87\% $\pm 2.61 \%$ | 5.29\% | 106.83\% $\pm 2.86 \%$ | 5.08\% |
| BCAA | 17.734 | 98.32\% $\pm 1.79 \%$ | 4.32\% | 99.79\% $\pm 2.45 \%$ | 5.05\% | 106.21\% $\pm 2.95 \%$ | 5.47\% |
| BDCAA | 19.815 | 100.96\% $\pm 2.66 \%$ | 5.03\% | 100.41\% $\pm 3.33 \%$ | 6.56\% | 106.46\% $\pm 3.82 \%$ | 6.72\% |
| DBCAA | 18.732 | 103.77\% $\pm 2.21 \%$ | 4.17\% | 101.30\% $\pm 3.12 \%$ | 6.13\% | 107.19\% $\pm 2.94 \%$ | 5.19\% |
| DALAPON | 10.879 | 99.11\% $\pm 2.36 \%$ | 5.16\% | 101.74\% $\pm 1.41 \%$ | 3.63\% | 101.24\% $\pm 1.59 \%$ | 3.28\% |

## References:

Agilent 1200 Series, Reference Manuals.
Agilent Triple Quad LC/MS, Maintenance and Familiarization Guides.
Agilent 6400 QQQ LC/MS Techniques and Operation, Agilent Technologies Course Number R1893A, Student Manuals Volumes 1 and 2.

