

METHOD STATEMENT



Determinand:

Total coliforms and Escherichia coli.

Matrix:

Sample Types: Waters.

Principle of Method:

A known volume of water sample is filtered through a membrane filter with 0.45 µm pores upon which bacteria are entrapped. The filter is then placed on a selective growth medium m-Lactose Glucuronide Agar which contains lactose, phenol red as an indicator of acidity and the chromogenic substrate 5-bromo-4-chloro-3-indolyl-b-D-glucuronide (BCIG) to indicate the production of b-glucuronidase.

Plates are incubated at 30°C for 4 hours and at 37°C for a further 14 hours after which colonies characteristic of coliforms and Escherichia coli are counted and picked off for confirmation where necessary.

Presumptive total coliforms appear as yellow, green or blue colonies. Presumptive Escherichia coli appear as green colonies. Confirmed total coliforms produce b-galactosidase and are oxidase-negative. Confirmed Escherichia coli produce b-galactosidase, are oxidase negative and produce indole from nutrient agar supplemented with 1% tryptone within 24 hours at 44°C.

As an alternative to traditional confirmation, colonies may be identified directly using a MALDI-TOF MS system to perform protein profiling.

Sampling and Sample Preparation:

Once taken, microbiological samples should be transferred immediately to dark storage conditions and kept at a temperature between $5 \pm 3^\circ\text{C}$ for transport to the laboratory. If samples are not analysed immediately on receipt in the laboratory, they should be kept at a temperature between $5 \pm 3^\circ\text{C}$, in dark conditions until analysis commences.

Samples should be analysed as soon as practicable on the day of collection. In exceptional circumstances, if there is a delay, storage under the above conditions should not exceed 24 hours before the commencement of analysis.

Where an exceedance has occurred the customer should be informed or a statement reflecting this should be included with the report (except where the customer has been already made aware that this is occurring on a regular basis and requests this not to be applied).

Interferences

Chlorine and chloramines. Neutralise by adding sodium thiosulphate which at a concentration of 18mg/l should counteract up to 5mg/l of free and combined residual chlorine.

Process waters may contain different biocides and the use of sodium thiosulphate may not be appropriate under these conditions. Customers should provide guidance when obtaining quotes.

Performance of Method:

Limit of Detection:

The Limit of Detection for this method is calculated as detailed in internal Procedure GOP7.2B.

Uncertainty of measurement:

The Uncertainty of Measurement for this method is calculated as detailed in internal Procedure GOP7.6C and the results are recorded on GQF7.6.3.

References:

METHOD STATEMENT



DoE (Department of the Environment) (1989) Guidance on Safeguarding the Quality of Public Water Supplies. Department of the Environment, HMSO, London.

DoE (Department of the Environment) (1994) The Microbiology of Water 1994 Part 1 - Drinking Water. Report on Public Health and Medical Subjects No 71. Methods for the Examination of Waters and Related Materials. Department of the Environment, HMSO, London. ISBN 011-753010 7.

SARTORY, D.P. and HOWARD, L. (1992). A medium detecting beta-glucuronidase for the simultaneous membrane filtration enumeration of Escherichia coli and coliforms from drinking water. Letters in Applied Microbiology, 15, 273-276.

Development of New Method for Simultaneous Enumeration of Coliforms and E.coli from Potable Water. Severn Trent Water Quality Assurance Project Report No: WS142 by David Sartory. January 1992.

Comparison of coliform and E.coli counts on broth and agar forms of membrane lauryl sulphate medium. by D.P. Sartory et al. Shelton Laboratory and A.S. Faulkner et al Barnhurst Laboratory. Severn Trent Water Limited.

Supplement to the report by Sartory, et al, on Comparison of coliform and E.coli counts on broth and agar forms of membrane lauryl sulphate medium by P. Holmes et al. Wanlip Laboratory, Severn Trent Water Limited. June 1991.

Supplement to report by Sartory et al on Comparison of coliform and E.coli L.Niccols et al Mythe laboratory and R. Barthakur et al Warwick laboratory, Severn Trent Water Limited.

Validation of counting on MLGA plates Dr D Hard and E Parker December 2000.

Environment Agency - The Microbiology of Drinking Water (2002) - Part 4 - Methods for the Isolation and Enumeration of Coliform Bacteria and Escherichia Coli (including E. Coli 0157:H7). Manufacturers Instruction Manual; BBL CRYSTAL Identification Systems Enteric/Nonfermenter ID Kit.

Environment Agency - The Microbiology of Drinking Water (2002) - Part 2 - Practices and Procedures for Sampling.

Collins and Lyne's Microbiological Methods. Sixth Edition 1989. Page 133, Membrane Filter Counts.

Collins and Lyne's Microbiological Methods Sixth Edition 1989. Page 104, 105 (Oxidase test).

Laboratory Methods in Food and Dairy Microbiology. Harrigan and McCance 1976. Page 79.

BBL Dryslide Oxidase Manufacturers Instructions Ref. L-000/46, Revised May 1999.

Supplement evaluation of the performance of MLGA as compared to MLSB, STL Microbiology, July-Dec 1997